

Growth performance, carcass traits, and blood biochemistry of broiler chicks fed with low-fibre sunflower meal and phytase

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(Received 11 March 2019; Accepted 04 June 2019; First published online 13 August 2019)

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Abstract

This study was designed to evaluate the impact of graded replacements of soybean meal (SBM) with low-fibre sunflower meal (LF-SFM), with and without phytase supplementation, on growth performance, carcass traits, and plasma profile of broilers. A total of 800 mixed sex one-day-old broiler chicks (Cobb 500) were randomly assigned to eight dietary treatment groups (five replicates each) in a 4 × 2 factorial arrangement, including four levels of LF-SFM (0%, 25%, 50%, and 75% to replace SBM) and two levels of microbial phytase (0 or 0.2 g/kg diet). Feed conversion ratio (FCR), and bodyweight gain (BWG) were decreased significantly if LF-SFM replaced more than 25% SBM. There was no significant effect between the interaction of the main factors (LF-SFM × phytase) on growth performance during the starter, grower, finisher and the overall period. The addition of phytase had no beneficial effects on performance traits. Carcass traits were not influenced by feeding LF-SFM or enzyme addition. However, increasing the level of LF-SFM (50% and 75%, respectively) in diets reduced the abdominal fat, whereas the weight of the small intestine was increased. Plasma metabolic profile was not affected by LF-SFM levels in broiler diets, except for high-density lipoproteins cholesterol (HDL-C) and total cholesterol (T-Cho), which were not affected by the dietary enzyme. It is concluded that the diets with LF-SFM levels up to 25% generated growth performance and carcass traits comparable with the diet that contained SBM.

Keywords: Cobb 500, digestive organs, enzyme, growth traits, low-fibre sunflower meal, plasma profile

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Introduction

The utilization of soybean protein in human food and animal feeds may cause problems in the future, such as availability, production costs, and the risks associated with reliance on a single ingredient. Soybean meal (SBM) is the main protein source in poultry feed, and is usually known for its high quality (NRC, 1994). SBM comes mostly from genetically modified crops. This situation encourages European Union (EU) countries to find ways to reduce dependence on imported protein feeds, which would increase the security of feed supply (Ciurescu *et al.*, 2017). Utilizing agro-industry by-products as inexpensive alternatives in poultry nutrition is of ongoing interest for nutritionists and poultry producers.

Sunflower (*Helianthus annuus* L.) is a high oil-yielding seed crop that is cultivated worldwide, and adapts well to a wide range of climatic and soil conditions. Sunflower meal (SFM) is an important by-product of sunflower oil extraction, which is a source of vegetable protein and fibre for human beings and animals (Kalmendal *et al.*, 2011). Its protein content depends on dehulling, air-classification, and oil extraction processes (Laudadio *et al.*, 2013). Additionally, SFM contains low antinutritional compounds, such as chlorogenic acid (Canibe *et al.*, 1999), and is more resistant to contaminants, compared with SBM (Bozzo *et al.*, 2008). Nevertheless, the use of SFM in poultry diet is limited by variations in its chemical composition. The two main components that restrict its use are its high fibre and low lysine contents (Rezaei & Hafezian, 2007; de Morais Oliveira *et al.*, 2016). One of these issues may be overcome by reducing the fibre content of SFM. The application of micronization, coupled with air fractionation, was considered a useful tool for improving some technological performances and nutritional properties and enriching the meal fractions of

healthy compounds (Rizzello *et al.*, 2012). High SFM levels in broiler diets require the addition of high levels of oil to compensate for the low energy content of this ingredient. Nevertheless, according to current market prices, oil seems to be one of the most expensive feedstuffs in broiler diets.

Phosphorus (P) is the third most expensive nutrient in poultry diets after energy and protein (Biehl *et al.*, 1998). Phytic acid (*myo*-inositol hexaphosphate) is an important plant P storage form and accounts for 50% to 80% of total P in plant seeds that are commonly used in livestock animal feeds. However, phytate-P has low bioavailability and is underutilized owing to the lack of endogenous phytate degrading enzymes in nonruminant livestock, including poultry (Selle *et al.*, 2006).

Phytase (*myo*-inositol-exakisphos-phohydrolase), a specific phosphohydrolase, degrades phytate to yield inositol monophosphate and orthophosphate via inositol penta- to monophosphates as intermediary products (Liu *et al.*, 1998). In-feed administration of microbial phytases to improve the digestibility of phytic acid is widely used in the production of poultry and other livestock (Simons *et al.*, 1990; Selle *et al.*, 2007; Cowieson *et al.*, 2011). Promising results have been reported when some exogenous enzyme is added to chicken, guinea fowl and quail diets that contain SFM to aid fibre digestion (carbohydrases) or to solubilize phytate-P (phytase), thereby reducing their negative effects on broiler production parameters (Mandal *et al.*, 2005). However, no research appears to have been reported on the application of phytase in broiler diets containing low-fibre SFM (LF-SFM).

Thus, the objective of this study was to evaluate an appropriate inclusion level of LF-SFM in chicks' diet as an alternative protein source instead of SBM, with or without microbial phytase supplementation, on the productive performance, digestive organ size and plasma biochemistry of broilers.

Materials and Methods

The study protocol was approved by the Local Ethical Committee for Animal Experiments in Balotesti, and was in accordance with the principles of EU Directive 2010/63/EU and Romanian Law on Animal Protection.

One-day-old Cobb 500 broiler chickens ($n = 800$) of mixed sex were purchased from a local commercial hatchery, weighed (43.4 ± 0.23 g/chick) individually, and randomly distributed to one of the eight dietary treatments in a 42-day feeding trial. Each treatment was sub-divided into five replicates, which were reared in pens of equal size (1.9×1.0 m) and considered one block. Pens were arranged in longitudinal lines in the house. The chicks were vaccinated at hatch for Marek disease, infectious bronchitis, and Newcastle disease and were raised on wood shavings litter. Heat was provided with a heating lamp per pen. Except for day 1, a 23-hour light to 1-hour dark lighting programme was applied during the experiment. The temperature was maintained at 32 °C at placement, followed by a 3 °C decrease each week to achieve 20–21 °C by using thermostatically controlled heaters, fans, and adjustable sidewall inlets. The main diets were arranged in a 4×2 factorial design, with the variable being LF-SFM, which replaced SBM on four levels (0%, 25%, 50%, and 75%, respectively) and enzyme supplementation at two levels (0 and 0.2 g/kg diet, respectively). The microbial phytase (Axta® PHY 5000 L, a *Buttiauxella* spp. bacterium, expressed in a *Trichoderma reesei* fungus) was produced by Danisco Animal Nutrition, DuPont Industrial Biosciences (Marlborough, UK), at 1000 phytase units (FTU)/kg of final feed. One FTU is defined as the quantity of enzyme required to liberate $1 \mu\text{mol}$ of inorganic P/min, at pH 5.5, from an excess of $15 \mu\text{M}$ sodium phytate at 37 °C (International Union of Biochemistry and Molecular Biology, 1992). The commercial powdered enzyme was added at 200 g/to 1000 FTU/kg feed (on top), as recommended by the manufacturer. The chemical compositions of diets are presented in Table 1.

The feeding programme was divided into three feeding phases: starter (days 1–10), grower (days 11–22), and finisher (days 23–42). Diets for each feeding phase were formulated to be isocaloric, isonitrogenous, with similar content of total lysine, total sulfur amino acids (TSAA) (methionine + cysteine), calcium and available phosphorous, and to meet or exceed breeder guidelines (Cobb Vantress Europe Ltd, UK). Feed (in mash form) and water were provided ad libitum throughout the trial. Samples of ingredients and feed were analysed in duplicate for content of dry matter, crude protein ($N \times 6.25$), ether extract and ash, using standard procedures according to the methods of the Commission Regulation (EC) no. 152 (OJEU, 2009). The contents of neutral detergent fibre (NDF) and acid detergent fibre (ADF) were determined using a Fibertec apparatus (automatic system Foss Tecator, Höganäs, Sweden). Carbohydrate content was estimated as nitrogen-free extract (NFE). Amino acids (AA) (excluding tryptophan, which was not determined) were analysed using a Thermo Scientific Surveyor Plus HPLC System (Thermo Fisher Scientific, MA, USA), according to the conditions described by Ciurescu & Pană (2017). Data are from the analysis carried out by the chemistry laboratory of National Research Development Institute for Biology and Animal Nutrition in Balotesti, Romania.

Table 1 Ingredients and nutrient composition (as-dry basis) of diets used in the study

Diets	Low-fibre SFM levels as a substitute for SBM (%)											
	Starter diets (0–10 days)				Grower diets (11–22 days)				Finisher diets (23–42 days)			
	0	25	50	75	0	25	50	75	0	25	50	75
<i>Ingredients (%)</i>												
Corn	57.62	58.64	58.62	58.76	67.43	68.24	67.94	67.68	68.54	69.05	68.72	68.41
Soybean meal (CP 46%)	31.40	22.50	15.00	7.50	22.0	15.00	10.00	5.00	21.50	15.00	10.00	5.00
LF-SFM (CP 44%)	0.0	7.50	15.00	22.50	0.0	5.00	10.00	15.00	0.0	5.00	10.00	15.00
Corn gluten meal (CP 62%)	4.00	4.60	4.50	4.30	4.00	5.60	6.00	6.20	3.00	4.20	4.50	4.80
Sunflower oil	2.00	1.70	1.70	1.70	1.50	1.10	1.00	1.00	2.50	2.20	2.20	2.20
Monocalcium phosphate	1.67	1.56	1.5,3	1.45	1.56	1.50	1.45	1.42	1.38	1.34	1.28	1.25
Calcium carbonate	1.45	1.46	1.4,6	1.46	1.39	1.42	1.42	1.42	1.25	1.28	1.29	1.27
Salt	0.28	0.28	0.2,8	0.28	0.28	0.28	0.28	0.28	0.28	0.28	0.28	0.28
L-Lysine (78%)	0.25	0.45	0.60	0.75	0.42	0.56	0.63	0.72	0.27	0.39	0.48	0.55
DL-Methionine (99%)	0.25	0.23	0.23	0.22	0.24	0.22	0.20	0.20	0.20	0.18	0.17	0.16
Choline chloride	0.08	0.08	0.08	0.08	0.08	0.08	0.08	0.08	0.08	0.08	0.08	0.08
Premix vit-min ¹	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0
Phytase ²	-/+ ³	-/+	-/+	-/+	-/+	-/+	-/+	-/+	-/+	-/+	-/+	-/+
<i>Calculated composition</i>												
ME (MJ/kg)	12.69	12.69	12.69	12.70	12.99	13.00	12.98	12.99	13.28	13.28	13.29	13.30
Crude protein	22.25	22.24	22.31	22.32	19.02	19.03	19.08	19.04	18.06	18.04	18.06	18.05
Lysine, total	1.30	1.30	1.30	1.30	1.19	1.19	1.19	1.19	1.05	1.05	1.05	1.05
Lysine, digestible	1.21	1.17	1.16	1.17	1.11	1.11	1.10	1.10	1.00	0.99	0.99	0.99
TSAA	0.98	0.97	0.98	0.97	0.89	0.89	0.88	0.89	0.82	0.82	0.82	0.82
Calcium	0.90	0.90	0.90	0.90	0.84	0.84	0.84	0.84	0.76	0.76	0.76	0.76
Available phosphorous	0.45	0.44	0.45	0.45	0.42	0.42	0.42	0.42	0.38	0.38	0.38	0.38
Crude fat	4.92	4.76	4.89	4.97	4.67	4.40	4.37	4.43	5.67	5.48	5.54	5.61
Crude fibre	2.80	3.30	3.84	4.38	2.53	3.07	3.67	4.27	2.51	3.06	3.66	4.26
<i>Analysed composition</i>												
Dry matter	88.2	87.6	88.7	89.2	86.5	87.9	88.6	89.0	86.2	87.4	87.7	87.6
Crude protein	22.12	22.21	22.77	22.65	19.27	19.35	19.42	19.33	18.32	18.39	18.45	18.43
Crude fat	4.88	4.75	4.82	4.94	4.78	4.25	4.23	4.30	5.56	5.34	5.69	5.72
Crude fibre	3.10	3.87	3.93	4.77	2.76	3.45	3.92	4.53	2.67	3.41	3.88	4.37

¹ Supplied per kg diet: 4.47 mg retinyl acetate, 0.12 mg cholecalciferol, 80 mg DL- α -tocopheryl acetate, 4 mg menadione sodium bisulphite, 4 mg thiamine mononitrate, 9 mg riboflavin, 4 mg pyridoxine-HCl, 0.020 mg cyanocobalamin, 15 mg Ca-pantothenate, 60 mg niacin, 2 mg folic acid, 100 mg Mn, 100 mg Zn, 40 mg Fe, 15 mg Cu, 1.0 mg I, 0.30 mg Se, 0.25 mg Co.

² Phytase added to diets at 0 or 0.2 g/kg (1000 FTU/kg feed)

³ - = enzyme not included in the diet; + = enzyme included in the diet

ME: metabolizable energy; TSAA: total sulfur amino acids

On days 11, 23, and 42, the birds and residual feed were weighed using a Kern scale (PCB 6000-0, Balingen, Germany) and the feed intake (FI), body weight gain (BWG), and feed conversion ratio (FCR) were calculated. Dead birds were weighed and removed, and the percentage mortality was calculated. At days 42 old, three broiler chickens per replicate pen (nearest to the average weight of the same pen) were selected, fasted for three hours with water being provided ad libitum, and killed by severing the right carotid artery and jugular vein. Slaughtered birds were weighed and their internal organs, including gizzard, liver, pancreas, and spleen, were removed, cleaned of adhering tissues, and weighed individually. The small intestines (duodenum, jejunum, and ileum) were also weighed and recorded. The cecum was quickly dissected, and the fresh contents were gently squeezed and carefully collected in 25 ml sterilized tubes. The cecal digesta

pH was measured (mean of three readings) with a portable pH-meter (WTW pH 3310, WTW GmbH, Weilheim, Germany). Carcasses, abdominal fat, and digestive organs were weighed, excluding head, neck, and feet. Carcass yield and relative weights of digestive organs were expressed as percentages of live bodyweight.

At day 42, blood samples were collected via the brachial vein of two broiler chickens per replicate pen, chosen randomly, into sterilized tubes (Vacutest Kima, 4ml, Italy) with heparin as an anticoagulant. The tubes were centrifuged at 3000 rpm for 15 minutes (Fuge D06, Neuation Technologies Pvt. Ltd, India). The resultant plasma was transferred to Eppendorf tubes and stored at -20 °C until further analyses. Total protein (T-Pro), albumin (Alb), total bilirubin (T-Bil), creatinine (Cre), urea (BUN), uric acid (UA), total cholesterol (T-Cho), triglycerides (TG), high-density lipoproteins cholesterol (HDL-C), calcium (Ca), magnesium (Mg), inorganic phosphorus (IP), alanine aminotransferase (ALT/GPT), aspartate aminotransferase (AST/GOT) and gamma-glutamyl-transferase (GGT) were determined with an automated biochemical analyser (dry chemistry system Spotchem EZ, SP- 4430 model; ARKRAY Global Business Inc., Kyoto, Japan) using the respective Arkray Diagnostic Kits.

Data were analysed with the general linear model (GLM) procedure of SPSS 20.0 software (2011). The statistical model included the effects of LF-SFM level, phytase addition, and their interaction. For growth performance (BWG, FI, and FCR), each pen was regarded as the experimental unit ($n = 5$). For other variables (carcass traits and plasma profiles) each sample was considered the experimental unit. Effects were considered statistically significant at $P < 0.05$. Probability values between 0.05 and 0.10 were considered trends. Post hoc multiple comparisons were performed by Tukey's test.

Results and discussion

The nutrient composition and amino acids profile of LF-SFM that were used for this study are shown in Tables 2 and 3. LF-SFM was found to contain an appreciable amount of crude protein compared with SBM (44–48.5%) as recorded by NRC (1994). In comparison with SBM, LF-SFM had lower metabolizable energy (7.10 vs 9.33 MJ) and threefold higher fibre content (11.31 vs 3.9 %).

Table 2 Nutrient composition and metabolizable energy of low-fibre sunflower meal, as fed basis ($n=2$)

Nutrients	DM	CP	EE	Fibre	Ash	NFE	NDF	ADF	ME (MJ)	Ca	P
LF-SFM	96.39	43.36	0.68	11.31	7.96	33.08	24.18	12.71	7.10	0.44	1.59

LF-SFM: low-fibre sunflower meal; DM: dry matter; CP: crude protein; EE: ether extract; NFE: nitrogen free extract; NDF: neutral detergent fibre; ADF: acid detergent fibre; ME: metabolizable energy; Ca: calcium; P: phosphorus

The metabolizable energy (ME) content of LF-SFM was estimated based on the following equation (NRC, 1994):

$$ME = 6.28 (DM) - 6.28 (\text{ash}) + 25.38 (CP) + 62.62 (EE),$$

Where: DM, CP, and EE are dry matter, crude protein, and ether extract, percentage of SFM, respectively.

All essential AA, except the lysine (3.7–4 % of the protein) were present in excessive amounts. LF-SFM had higher sulfur-containing amino acids (methionine and cysteine 2%), compared with the requirements of broilers in the starter phase.

Table 3 Amino acid profile (g/100 g) of low-fibre sunflower meal

AA	LF-SFM	AA	LF-SFM
Lysine	1.636	Phenylalanine	2.359
Methionine	0.715	Tyrosine	1.192
Cystine	0.659	Serine	2.592
Threonine	2.923	Glycine	2.650
Leucine	3.322	Alanine	2.326

Arginine	4.077	Aspartic acid	4.701
Isoleucine	1.905	Glutamic acid	9.991
Valine	1.958		

AA: amino acids; LF-SFM: low-fibre sunflower meal

The effects of LF-SFM level, phytase supplementation and interaction between level and enzyme on the performance of broiler chicks are presented in Table 4. Mortality was low (<2.5%) and unrelated to treatment. There was no significant effect ($P > 0.05$) between the interaction of the main factors (LF-SFM \times phytase) on BWG, FI, and FCR of broiler during the starter, grower and finisher phases and the overall period. In the first period of the trial (days 1-10), the results revealed that the broilers fed diets containing LF-SFM at 25% or 50% instead of SBM had comparable BWG. However, increasing LF-SFM in diets from 50 to 75% significantly ($P < 0.001$) decreased BWG.

FI and FCR were not significantly ($P > 0.05$) affected by the various LF-SFM inclusion levels. During the grower period (days 11–22), the diets with LF-SFM levels up to 25% generated BWG that was comparable with a diet containing SBM. Conversely, higher levels of LF-SFM (50% and 75%) recorded the lowest value of BWG ($P < 0.001$) and consequently impaired FCR ($P = 0.043$) in comparison with other levels. During the finisher phase (days 21–42), high inclusion levels of SFM (50% and 75%) posed the same problems, so resulted in a reduction of BWG ($P < 0.001$) and impaired FCR ($P = 0.023$), but had no effect on FI ($P > 0.05$). The addition of phytase in the diets had no impact on FI ($P > 0.05$), and tended ($P = 0.067$) to impair BWG and FCR ($P = 0.075$). Over the entire feeding period the poorest values of BWG ($P < 0.001$) and FCR ($P = 0.004$) were observed in chicks fed diets that contained 50% or 75% LF-SFM, whether supplemented with the enzyme or not. The addition of 0.2 g microbial phytase/kg diets was not effective in improving BWG, FCR, or FI ($P > 0.05$).

There are no reports about the use of LF-SFM in broiler chicken diets. Some studies are related to other poultry species. Most of them have shown that up to 15% and 18% processed SFM have been used with success in laying hens (Laudadio *et al.*, 2014a) and turkey diets (Laudadio *et al.*, 2014), respectively. Earlier studies that investigated the effect of the use of SFM as a replacement for SBM showed inconsistent results. The findings of the current study are in line with the results of Mandal *et al.* (2006), Abdelrahman & Saleh (2007) and Peric *et al.* (2010), who found that the inclusion SFM in broiler diets decreased growth performance in terms of poor feed efficiency and growth rate. On the other hand, in other studies, the inclusion of SFM up to 20% (Tavernari *et al.*, 2008) and even higher levels (Rama Rao *et al.*, 2006; Mushtaq *et al.*, 2009) did not have adverse effects on BW or BWG. Senkoylu *et al.* (2006) did not observe any effect on broiler performance when up to 28% SFM was included in the feed. However, those authors used the residue of SFM from cold pressing (32.3% CP and 18.78% EE). In the same context, Alagawany *et al.* (2017) reported that increasing SFM level in the diet up to 50% to replace SBM improved performance traits (including BW, BWG, and FCR). This difference may be attributed to the quality or level of the SFM processing or variety of the birds.

There were no significant differences among the treatments because of the interaction effects between dietary LF-SFM level and phytase supplementation on growth performance (Table 4). Phytase addition had no beneficial effects on performance traits. The poorest values (1,670 g) of BWG and 2.03 (g feed/g gain) of FCR were achieved by broilers fed LF-SFM supplemented with exogenous phytase in the finishing period. This finding suggests that the use of *Trichoderma reesei*-derived phytase was not efficacious in releasing phytate P from LF-SFM. In this context, Adeola & Cowieson (2011), Ranvindrana (2013), and Dersjant-Li *et al.* (2015) noted that the response of chickens to a phytase might be affected by various factors, such as the substrate (nutrients and antinutrients in the ingredients used in feed formulations) and enzyme origin (fungal vs. bacterial vs. yeast), and the conditions in the gastrointestinal tract, such as moisture content, temperature and pH.

Table 4 Dietary low-fibre sunflower meal level and phytase addition on growth performance of broiler chickens

Treatments	LF-SFM (%)	Phytase ¹ (g/ kg diet)	1–10 days			11–22 days			23–42 days			1 to 42 days		
			BWG (g)	FI (g)	FCR (g/ g)	BWG (g)	FI (g)	FCR (g/ g)	BWG (g)	FI (g)	FCR (g/ g)	BWG (g)	FI (g)	FCR (g/ g)
1 ²	0	0	228	368	1.61	670	1,021	1.52	1,793	3,490	1.95	2,692	4,878	1.81
2	25	0	226	366	1.62	659	1,004	1.52	1,790	3,481	1.94	2,675	4,851	1.81
3	50	0	221	363	1.64	622	974	1.57	1,673	3,378	2.02	2,516	4,714	1.87
4	75	0	216	357	1.65	619	972	1.57	1,662	3,360	2.02	2,497	4,689	1.88
5	0	0.2	227	370	1.63	673	1,025	1.52	1,755	3,443	1.96	2,656	4,840	1.82
6	25	0.2	224	364	1.63	652	1,006	1.54	1,738	3,419	1.97	2,615	4,788	1.83
7	50	0.2	223	367	1.65	644	998	1.55	1,605	3,341	2.08	2,472	4,705	1.90
8	75	0.2	217	359	1.65	639	993	1.55	1,579	3,346	2.12	2,435	4,697	1.93
SEM			1.70	4.40	0.01	3.70	10.40	0.01	11.30	17.40	0.01	13.40	35.60	0.01
Main effects ³														
Level (L)														
0			228 ^a	369	1.62	672 ^a	1,024	1.52 ^b	1,774 ^a	3,466	1.96 ^b	2,674 ^a	4,858	1.82 ^b
25			225 ^{ab}	365	1.63	656 ^a	1,005	1.53 ^{ab}	1,764 ^a	3,450	1.96 ^b	2,645 ^a	4,820	1.82 ^b
50			222 ^{bc}	364	1.65	634 ^b	986	1.55 ^a	1,625 ^b	3,361	2.07 ^a	2,480 ^b	4,710	1.90 ^a
75			217 ^c	358	1.65	630 ^b	983	1.56 ^a	1,621 ^b	3,353	2.07 ^a	2,466 ^b	4,693	1.90 ^a
Phytase (PHY)														
No			224	364	1.63	644	993	1.54	1,730	3,427	1.98	2,595	4,783	1.85
Yes			223	365	1.64	652	1,006	1.54	1,670	3,388	2.03	2,538	4,759	1.88
P-value														
L effect			<0.0001	0.178	0.442	<0.0001	0.121	0.043	<0.0001	0.340	0.023	<0.0001	0.127	0.004
PHY effect			0.634	0.924	0.754	0.527	0.381	0.929	0.067 ^T	0.545	0.075 ^T	0.348	0.432	0.534
L x PHY effect			0.826	0.887	0.927	0.898	0.927	0.929	0.924	0.765	0.768	0.924	0.688	0.670

^{a,b,c} Columns means with different superscripts differ significantly at $P < 0.05$

¹Axtra® PHY 5000 L (1000 FTU/kg feed)

²Control group

³Data were analysed as 4 × 2 factorial arrangement
LF-SFM: low-fibre sunflower meal

Table 5 Means of dietary low-fibre sunflower meal level and phytase addition on carcass traits of broiler chickens¹

Treatments	LF-SFM (%)	Phytase ² (g/kg)	Carcass ⁵	Breast ⁶	Abdominal fat ⁶	Digestive organs weight ⁷					Cecal pH
						Gizzard	Liver	Pancreas	Spleen	Small intestine	
1 ³	0	0	72.9	29.2	1.57	2.19	2.60	0.30	0.09	3.03	6.56
2	25	0	72.4	28.7	1.46	2.16	2.53	0.29	0.11	3.10	6.62
3	50	0	71.8	28.6	1.28	2.22	2.62	0.25	0.10	3.17	6.59
4	75	0	72.6	28.9	1.22	2.28	2.59	0.31	0.11	3.21	6.64
5	0	0.2	71.5	28.4	1.55	2.14	2.47	0.28	0.09	3.09	6.55
6	25	0.2	70.9	28.8	1.42	2.17	2.59	0.29	0.10	2.90	6.50
7	50	0.2	70.7	28.5	1.31	2.25	2.61	0.30	0.08	3.11	6.60
8	75	0.2	71.4	28.3	1.24	2.17	2.52	0.26	0.11	3.14	6.57
SEM			1.10	0.62	0.09	0.24	0.19	0.04	0.01	0.33	0.22
Main effects ⁴											
Level (L)											
			72.2	28.8	1.56 ^a	2.17	2.54	0.29	0.09	3.06 ^b	6.55
			71.7	28.7	1.44 ^b	2.16	2.56	0.29	0.11	2.99 ^b	6.56
			71.4	28.7	1.29 ^{bc}	2.20	2.61	0.28	0.09	3.14 ^a	6.60
			72.0	28.6	1.23 ^c	2.23	2.56	0.29	0.11	3.18 ^a	6.61
Phytase (PHY)											
			72.4	28.8	1.38	2.21	2.59	0.29	0.10	3.13	6.60
			71.2	28.5	1.38	2.18	2.55	0.28	0.09	3.06	6.56
<i>P</i> -value											
			0.467	0.543	<0.001	0.096 ^T	0.215	0.494	0.510	<0.001	0.583
			0.332	0.657	0.767	0.443	0.384	0.786	0.578	0.334	0.625
			0.682	0.951	0.546	0.567	0.333	0.624	0.862	0.476	0.563

^{a,b,c} Columns means with different superscripts differ significantly at $P < 0.05$

¹Data are means of 15 broilers for each treatment

²Axtra® PHY 5000 L (1000 FTU/kg feed)

³Control group

⁴Data were analysed as 4 × 2 factorial arrangement

⁵Represents as weight (g) of without head, neck and feet carcass as 100 g of live body weight

⁶Calculated as weight (g) of carcass characteristics to 100 g of live body weight

⁷Represents as weight (g) of organ weights as 100 g of live body weight

Table 6 Means of dietary low-fibre sunflower meal levels and phytase addition on plasma metabolic profile of broiler chickens¹

Treat-ments	LF-SFM (%)	Phy ² (g/kg)	Energy profile				Protein profile				Mineral profile			Enzyme profile			
			T-Cho mg/dL	HDL-C U/L	TG mg/dL	T-Pro g/dL	Alb g/dL	T-Bil mg/dL	Cre mg/dL	UA mg/dL	BUN mg/dL	Ca mg/dL	IP mg/dL	Mg mg/dL	ALT/GPT IU/L	AST/GOT IU/L	GGT U/L
1 ³	0	0	115.2	97.8	55.2	2.04	0.78	0.12	0.56	4.62	3.20	12.74	6.60	1.86	9.4	278.4	47.8
2	25	0	133.0	119.2	47.0	2.34	0.84	0.16	0.58	5.06	2.00	12.96	6.82	1.98	9.4	332.4	46.0
3	50	0	109.0	83.6	84.4	2.14	0.68	0.16	0.58	4.14	2.00	12.36	6.80	1.98	8.6	272.6	42.8
4	75	0	111.4	82.4	78.0	2.28	0.82	0.14	0.60	4.74	2.00	12.66	6.84	2.02	7.6	234.6	46.0
5	0	0.2	108.4	89.8	31.8	2.08	0.76	0.14	0.64	4.88	2.00	12.24	6.24	2.12	7.6	236.6	39.2
6	25	0.2	131.6	131.8	55.8	2.34	0.98	0.16	0.60	5.08	2.00	12.72	7.18	2.02	10.4	322.8	46.8
7	50	0.2	122.0	110.2	52.4	2.30	0.80	0.18	0.58	4.58	2.00	12.90	6.82	2.08	7.0	273.6	42.6
8	75	0.2	122.4	115.2	76.6	2.18	0.74	0.14	0.66	4.68	2.00	13.18	7.68	2.06	7.6	301.4	46.6
SEM			5.56	8.57	13.89	0.12	0.07	0.03	0.03	0.56	0.26	0.44	0.30	0.09	1.45	57.98	3.31
Main effects ⁴																	
Level (L)																	
		0	111.80 ^b	93.80 ^b	43.5 ^T	2.06	0.77	0.13	0.60	4.75	2.60 ^T	12.49	6.42 ^T	1.99	8.5	257.5	43.5
		25	132.30 ^a	125.50 ^a	51.4	2.34	0.91 ^T	0.16	0.59	5.07	2.00	12.84	7.00	2.00	9.9	327.6	46.4
		50	115.50 ^b	96.90 ^b	68.4	2.22	0.74 ^T	0.17	0.58	4.36	2.00	12.63	6.81	2.03	7.8	273.1	42.7
		75	116.90 ^b	98.80 ^b	77.3 ^T	2.23	0.78	0.14	0.63	4.71	2.00	12.92	7.26 ^T	2.04	7.6	268.0	46.3
Phytase (PHY)																	
		No	117.15	95.75 ^b	66.15	2.20	0.78	0.15	0.58	4.64	2.30	12.68	6.76	1.96	8.75	279.5	45.65
		Yes	121.10	111.75 ^a	54.15	2.22	0.82	0.16	0.62	4.80	2.00	12.76	6.98	2.07 ^T	8.15	283.6	43.80
<i>P</i> -value																	
		L effect	0.004	0.003	0.080 ^T	0.171	0.096 ^T	0.429	0.532	0.661	0.064 ^T	0.750	0.057 ^T	0.932	0.390	0.629	0.582
		PHY effect	0.323	0.013	0.231	0.774	0.426	0.648	0.119	0.680	0.112	0.797	0.319	0.088 ^T	0.561	0.921	0.435
		L x PHY effect	0.237	0.104	0.439	0.763	0.339	0.974	0.663	0.969	0.064 ^T	0.536	0.249	0.565	0.735	0.819	0.437

^{a,b,c} Columns means with different superscripts differ significantly at $P < 0.05$

¹Data are means of 10 broilers for each treatment; ²Axtra® PHY 5000 L (1000 FTU/kg feed); ³Control group; ⁴Data were analysed as 4 × 2 factorial arrangement
T-Cho: total cholesterol; HDL-C: high-density lipoproteins cholesterol; TG: triglycerides; T-Pro: total protein; Alb: albumin; T-Bil: total bilirubin; Cre: creatinine; UA: uric acid; BUN: urea; Ca: calcium; IP: inorganic phosphorus; Mg: magnesium; ALT/GPT: alanine aminotransferase; AST/GOT: aspartate aminotransferase; GGT: gamma-glutamyl-transferase

On the other hand, Leske & Coon (1999) reported that phytates from different ingredients are not similarly susceptible to dephosphorylation and that the reactive phytate content, and not total, is critical in determining the responses to supplemental phytase. They also found for example that canola meal contained a relatively high level of total phytate, but less reactive phytate, and did not respond well to added phytase. On the contrary, Iyayi *et al.* (2013) showed that microbial phytase (*Aspergillus oryzae*; 1000 FTU/kg) could have beneficial effects on black-eyed pea and peanut flour diets in broiler chickens. Furthermore, Cowieson & Adeola (2005) and Juanpere *et al.* (2005) found that phytase in combination with carbohydrase and protease had additive effects in nutritionally marginal broiler diets.

The effects of dietary LF-SFM and microbial phytase inclusion or interaction between level and enzyme on carcass characteristics, internal organ weight and pH of the cecal digesta of broiler chickens (at day 42) are summarized in Table 5. The interaction between the main factors (LF-SFM × phytase) had no significant effects ($P > 0.05$) on carcass yield, breast yield, abdominal fat, and digestive organ weight or cecal pH of broilers. However, increasing the level of LF-SFM (50% and 75%) in diets reduced abdominal fat ($P < 0.001$), whereas, the weight of small intestine was increased ($P < 0.001$) and tended ($P = 0.096$) to increase gizzard weight. Broilers fed diets that contained phytase or did not had no effect on carcass yield, breast yield, abdominal fat and digestive organ weight (*i.e.* gizzard, liver, pancreas, and spleen) and cecal pH ($P > 0.05$). Laudadio *et al.* (2014b) found that the substitution of SBM with micronized air-classified SFM (172 g/kg) as the main protein source had no adverse effect on growth performance and dressing percentage, and improved the breast and thigh muscle relative weights of the turkeys. Alagawany *et al.* (2017) observed that increasing dietary inclusion of sunflower meal (0%, 25%, 50%, and 75% replacing SBM) in broiler diets negatively influenced abdominal fat percentage. Araújo *et al.* (2011) reported that use of sunflower meal up to 15% in the broiler diets did not influence growth performance and carcass and yield from 22 to 42 days old. On the contrary, the inclusion of sunflower meal (0%, 8%, 16%, and 24%) in chick diets negatively influenced performance and carcass parameters (Araújo *et al.*, 2014). Brenes *et al.* (2008) found that including 250 g/kg of sunflower seeds in broiler diets caused a negative effect on relative liver weight and relative duodenum, jejunum, ileum and ceca lengths. In the current study, the increased weights of the small intestine when feeding diets with SFM (50% and 75%) to replace SBM for the entire rearing period could be the result of dietary fibre (DF). In general, DF ingestion leads to increased size and length of the digestive organs, including the small intestine, colon, and caecum of broilers (Iji *et al.*, 2001). These effects are often associated with modification of the gut epithelium morphology, and consequently with the hydrolytic and absorptive functions of the epithelium. Ciurescu *et al.* (2017) also observed an increase in the relative weight of the small intestine in chickens fed with legumes beans, such as lentils (*Lens culinaris*; cv. Eston, green seeded and cv. Anicia, green marbled seeded).

The plasma metabolic profile of broilers is listed in Table 6. There was no interaction effect between the main factors (LF-SFM × phytase) on plasma protein, energy, mineral and enzymatic profiles, but data showed that the level of LF-SFM tended to decrease plasma BUN ($P = 0.064$). Increasing LF-SFM up to 75% to replace SBM increased plasma T-Cho ($P = 0.004$) and HDL-C ($P = 0.003$) significantly, and tended to increase TG ($P = 0.080$), Alb ($P = 0.096$), BUN ($P = 0.064$) and plasma IP concentration ($P = 0.057$), compared with control (broilers fed SBM). The inclusion of microbial phytase in the LF-SFM diets increased plasma HDL-C ($P = 0.013$) and tended to increase Mg ($P = 0.088$) concentration. Blood biochemical data are commonly used as a diagnostic tool in poultry veterinary medicine, because they can reflect the condition of the organism and the changes happening to it under the influence of nutritional and environmental factors. Thus, clinical biochemical analysis can detect metabolic disorders and subclinical diseases that affect the production efficiency. The current results agree partially with those of Rama Rao *et al.* (2006), who reported that increased content of SFM ($\geq 67\%$ of SBM) in the broiler diets had increased the concentration of HDL-C, while the concentration of low-density lipoproteins cholesterol (LDL-C) decreased. They also noticed that the serum protein and triglycerides concentration decreased progressively with the increase in dietary SFM. It is well known that higher DF content reduces dietary fat utilization by deconjugating bile salts (Story & Furumoto, 1990), which might have reduced fat absorption through the gut. Thus, the body fat and, in particular the fat of the liver, could be used for metabolic needs and for increasing HDL serum concentration. On the other hand, Brenes *et al.* (2008) reported that the inclusion of 250 g/kg of sunflower seeds in the diets of broiler chicks had a negative impact on blood parameters. Alagawany *et al.* (2017) also indicated that using SFM at 0%, 25%, 50%, and 75% instead of SBM increased concentrations of total protein and its fractions (albumin and globulin). In contrast, Adejumo & Williams (2006) observed that total protein, albumin, and globulin were not affected by dietary SBM being replaced by SFM at 0%, 25%, 50%, and 75%, during the growing period. In the present study, there were no significant differences ($P > 0.05$) in the activity of ALT/GPT, AST/GOT, and GGT between treatments, that is, enzymes that are frequently used in evaluating hepatic function. Neither LF-SFM nor phytase addition had any effect on liver health. Previously, Alagawany *et al.* (2017) reported that the activity of ALT was enhanced with increasing the SFM level up to 50% in

broiler diets. To the best of the authors' knowledge, there are no published studies that test the specificity between LF-SFM and *Buttiauxella*-derived phytase on broiler blood constituents. Viveros *et al.* (2002) showed that the addition of phytase (Natuphos 500) in broiler low-P diets increased plasma P level and serum AST activity, reduced plasma Ca and Mg contents, and reduced serum ALT, and alkaline phosphatase, and lactate dehydrogenase activities.

Conclusion

The replacement of SBM in broiler chicken diets with LF-SFM at levels lower than 25% resulted in similar productive performance, carcass and breast yield, digestive organ size and pH of the cecal digesta. Higher LF-SFM levels (50% and 75%) had a negative effect on small intestine weight, and tended to increase gizzard weight. Phytase addition had no favourable effect on BWG, FI, and FCR. The health state of the broilers was not affected. Further research about the practical implications of microbial phytase is recommended before it is applied on a commercial scale.

Acknowledgements

This study was supported financially by the Ministry of Research and Innovation of Romania, and the UEFICDI as research funding agency (Grant No. 17 PFE/ 17.10.2018, Grant No. 2018-PCCDI-8/2 and Grant No. PN 19 09 01 04).

Authors' Contributions

GC, AV and HG contributed to experimental design, data analysis, and interpretation. GC and DG contributed to experiment care, sample processing, and data recording. GC and AV contributed to manuscript writing and manuscript preparation.

Conflict of Interest Declaration

The authors declared no conflict of interest.

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